
Role of Maternal Serum Ferritin in the Prediction of Asymmetric Intrauterine Growth Restriction.

Short running title: Ferritin predicts asymmetric IUGR

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Disclosure statement

The authors guarantee that they:

- (1) have stated explicitly, in the cover letter and the main text, that there are no conflicts of interest in connection with this article.
- (2) have participated sufficiently in the work to take responsibility for it;
- (3) have self-funded this research;
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All of the included women gave their **informed consent** prior to their inclusion in the study and their anonymity was maintained.

Abstract

Background and aim: Asymmetric intrauterine growth restriction (IUGR) carries increased intrauterine and neonatal risks. Since most cases are idiopathic, identifying a predictive test remains an ideal prevention management. Maternal serum ferritin level is a potential predictor of later developing asymmetric IUGR.

Methods: Blood samples were drawn, at 30-32 gestational weeks, from 450 women who were then followed-up, resulting in 32 term pregnancies with asymmetric IUGR. The control group included the first recruited 32 women delivering adequate for gestational age neonates at term. Serum ferritin was then measured in the stored serum samples. Ultrasound scanning was performed at 30-32 weeks then at 37 weeks. Umbilical and middle cerebral artery Doppler scans were added at 37 weeks.

Results: Serum ferritin, at 30-32 weeks, was higher in women delivering IUGR babies (19.3 ± 6.83 vs 14 ± 5.18 , $p < 0.01$). At 37 weeks, pregnancies with asymmetric IUGR had significantly ($p < 0.01$) higher umbilical artery resistance and pulsatility indices (0.71 ± 0.06 vs 0.59 ± 0.07 and 1.24 ± 0.17 vs 0.86 ± 0.09 , respectively). Their middle cerebral artery RI and PI were significantly ($p < 0.01$) lower than

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controls (0.74 ± 0.09 vs 0.81 ± 0.05 and 1.64 ± 0.07 vs 1.9 ± 0.15 , respectively). Ferritin level correlated significantly with neonatal birthweight. A cut-off level >18.2 ng/mL had a sensitivity of 59.4% and a specificity of 90.6%. The area under curve showed an accuracy of 76.8%. Women with ferritin >18.2 ng/ml, were 10.23 times more likely to get asymmetrically growth restricted neonates (CI 2.89–36.17, $p<0.001$).

Conclusion: This study presents further evidence that maternal serum ferritin may be a useful test in the prediction of asymmetric IUGR.

Key words:

Intrauterine Growth Restriction,
Doppler,
Ferritin,
Neonatal Birthweight,
Small for gestational age.

Synopsis

Maternal serum ferritin may be a useful test in the prediction of asymmetric IUGR.

Introduction

Asymmetric, late-onset (type II), intrauterine growth restriction (IUGR) may be defined as pathological slow fetal growth, beginning in late pregnancy, due to uteroplacental insufficiency (1,2). This definition differentiates it from the term “Small for Gestational Age” (SGA) which implies a birthweight less than the 10th centile for gestational age even if it is a healthy but simply small baby (2). The incidence of IUGR is 3.3–10% in the developed countries and 6.7–17% in developing ones (3,4,5).

A fetus with IUGR is exposed to increased intrauterine risks of fetal distress and death, neurologic developmental disorders as well as meconium aspiration at birth. Neonatal risks include hypoglycemia, long admission to intensive care units, hypothermia, polycythemia, jaundice, feeding difficulties, necrotizing enterocolitis, late-onset sepsis, hypoxic-ischemic encephalopathy and pulmonary hemorrhage. These infants also have increased risks of type 2 diabetes, obesity, autoimmune diseases, cardiovascular diseases and hypertension in adult life (2,6).

Asymmetric IUGR, accounting for 70–80% of cases of IUGR (2), is associated with reduced umbilical blood flow, brain sparing effect, oligohydramnios and a low Ponderal index (7). In the absence of any direct causal therapy, and in view that most cases of IUGR are idiopathic, the identification of adequate predictive tests for IUGR remains one of the top priorities in obstetrics. Several studies suggested different biomolecules as early markers for IUGR, such as lactate dehydrogenase, leptin, metastin, adiponectin, s-endoglin, endothelin-1 and pregnancy associated plasma protein (8–10). Most of these tests have a low sensitivity, or are expensive, not widely available and/or invasive tests requiring amniocentesis (9,10).

Ferritin, the main iron storage protein, was suggested to be an adequate alternative as a screening test, being a relatively cheap and easily available blood test. Its level is known to rise in response to hypoxia or as an acute phase reactant in infections (11). Ferritin blood level normally drops, in correlation with the progressive depletion of iron reserves, by 32% during the first trimester, 39% in the second and up to 53% during the third trimester (12). This level reaches a nadir at 30–32 weeks (13), after which it stays constant.

The theory behind the rise in maternal serum level of ferritin during pregnancies destined to develop late-onset (asymmetric) IUGR relies on decreased placental extraction of ferritin from the systemic circulation (13). Its predictive value was previously investigated, in a few small-sized studies, on a small number of cases (9,13–16).

This study aimed to assess the accuracy of maternal serum ferritin level at 30 weeks to 32 weeks in predicting the development of IUGR, and to identify the ferritin cutoff level with the best predictive value, in a well-selected, adequate-sized group of pregnant women.

Material and Methods

This was a prospective nested longitudinal study conducted in the outpatient clinics of Ain Shams University Maternity Hospital, Cairo, Egypt. The study protocol was approved by the Faculty of Medicine Research Review Board on 15/03/2016. The study protocol was registered on 14/04/2016 on www.clinicaltrials.com website as NCT02738463.

All women were informed about the procedures of the study. Confidentiality of the collected data and anonymity of the participants were ensured. Written informed consent was obtained from the women before participation. The study protocol complied with the Declaration of Helsinki regarding ethical conduct of research involving human subjects.

All included women had singleton 30-32 weeks pregnancies (with sure and reliable dates of the last menstrual periods and ultrasonographic estimation of the gestational age during the first trimester). Exclusion criteria included history of anemia, iron supplementation, recent blood transfusion, age <20 years, BMI <18, smoking, diabetes mellitus during pregnancy, hepatic, renal, hypertensive or cardiovascular abnormalities, detected congenital fetal anomalies, antepartum hemorrhage, preterm delivery, and cases developing acute infections during pregnancy with raised leukocytic count or C-reactive protein. This was meant to rule out any confounding effect on the level of serum ferritin.

Sample size calculation:

Data from a previous study showed that maternal serum ferritin levels in pregnancies resulting in neonates with IUGR were on average 6.43 mg/L higher than in those with adequate sized neonates (15). Using the equation of sample size calculation from Fox et al (17), with a power of 80% ($\beta=0.2$) and a confidence level of 95% ($\alpha=0.05$), the sample size calculation for the study group is as follows:

$$N = D \times K \times 2 \times \left[\frac{\sigma}{(f1 - f2)} \right]^2$$

$$= 133.3\% \times 7.9 \times 2 \times \left[\frac{7.84}{(17.54 - 11.11)} \right]^2$$

$$= 32 \text{ cases per group}$$

Where N= Sample size (in a single arm), K= Constant, which varies according to α and β values, here K= 7.9, σ = standard deviation in ferritin level, and f1 and f2= the ferritin level among cases versus controls in a previously published study (15). D= percentage increase to compensate for the drop-out rate, here D=133.3%.

Assuming an incidence of IUGR of 8% (15), 400 patients were to be recruited for follow up to obtain the desired 32 cases with IUGR. Another 10% rate was added to that number of women to ascertain obtaining the desired number of IUGR cases

during patient follow-up. Thus, a total of 450 patients were assessed and followed-up. The study group included patients delivering at ≥ 37 weeks, giving birth to neonates with type II IUGR (32 pregnancies). The control group included the first recruited 32 women delivering adequate for gestational age neonates at term.

Morning fasting blood samples were drawn from the cubital vein at 30-32 weeks. The specimens were centrifuged immediately for 5 minutes at 4000 rpm, and the supernatant serum was transferred into another Eppendorf tube and immediately frozen and stored at -20C in the biochemistry laboratory of Ain Shams University Hospitals.

Obstetric ultrasound scanning was performed on all included women at 30-32 weeks and was repeated at 37 weeks. Umbilical and middle cerebral artery Doppler scans were added at 37 weeks in IUGR cases (the first 32 women diagnosed with IUGR) as well as controls (the first 32 normal pregnancies who reached maturity).

Pregnant women were followed until delivery to identify those with asymmetric IUGR. Serum ferritin levels were then measured in their stored samples of cases and controls using Enzyme-Linked Fluorescent Assay. The result of their complete blood count at delivery was also recorded.

Neonates were then evaluated at birth for vitality using the Apgar score of 0-10 which checks the heart and respiratory rates, the muscle tone, reactivity and skin color. Neonatal weight and body length were also measured and documented, as well as any clinical evidence of IUGR, such as raised head to abdomen circumferences ratio, loose and dry skin, absent buccal fat (old man look), small or scaphoid abdomen, thin umbilical cord often stained with meconium, excessive skin folds with decreased underlying skeletal muscle mass and loss of subcutaneous fat.

Data were collected, revised, coded and entered to the Statistical Package for Social Science program (IBM Corp. Released 2016. IBM SPSS Statistics for Windows, Version 24. Armonk, NY: IBM Corp). Qualitative data were presented as number, and percentage, and were compared using Chi-square χ^2 test or Fisher exact test. Quantitative parametric data were presented as mean \pm standard deviation (\pm SD), and were compared using Inde-

pendent t-test. Non-parametric data were presented as median, with interquartile ranges (IQR), and were compared using Mann-Whitney test. Spearman correlation coefficients were used to assess the correlation between two quantitative parameters in the same group. Logistic regression analysis was used to assess predictors of IUGR. Receiver operating characteristic curve (ROC) was used to assess the best cut-off point for ferritin in predicting IUGR with its area under curve (AUC), sensitivity, specificity, positive predictive value (PPV) and negative predictive value (NPV).

Results

A total of 450 women were recruited during the course of this study. Four pregnancies ended with intrauterine death, ten women developed gestational diabetes mellitus, 28 developed preeclampsia, 32 delivered prematurely and 48 were lost to follow up. Out of the remaining 328 pregnancies, the first 32 cases of IUGR and 32 adequate for gestational age (AGA) controls were included in data analysis. The mean age of the included women were statistically similar (28 ± 4.1 and 28.4 ± 4 , in controls and cases, respectively, $p=0.7$). Both groups had comparable rates of previous IUGR deliveries (6.3 and 17% respectively, $p=0.17$).

Women with asymmetric IUGR had similar hemoglobin level, hematocrit value erythrocytic and leukocytic counts as controls (table 1). Serum ferritin at 30-32 weeks was significantly higher in women who ended up delivering IUGR babies (table 1). At the 37 weeks' Doppler scan, pregnancies with asymmetric IUGR had significantly higher umbilical artery resistance indices (RI) and pulsatility indices (PI). Their middle cerebral artery RI and PI were significantly lower than their controls (table 1).

Neonates with asymmetric IUGR had a similar mode of delivery and gestational age at birth as their AGA controls. On the other hand, the birth-weight, Apgar scores and rate of admission to neonatal intensive care units were all significantly different between IUGR and AGA neonates (table 2).

Maternal serum ferritin level was the only variable which correlated significantly with neonatal birth-weight. Maternal age and hematology variables, neonatal gestational age, Apgar scores or body

length were all not correlated to ferritin level (table 3).

There was a non-significant correlation between the history of IUGR in previous pregnancies and the development of asymmetric IUGR in the current pregnancy. A highly significant correlation was noted between serum ferritin level at 30-32 weeks as well as Doppler indices at 37 weeks and present history of asymmetric IUGR (table 4).

The receiver operating characteristic (ROC) curve (table 5, figure 1) showed that the best ferritin cut-off level, between mothers with asymmetric IUGR neonates and those with AGA, was >18.2 ng/mL. This cutoff had a sensitivity of 59.38% and a specificity of 90.62%. The area under curve (AUC) showed an accuracy of 76.8%, thus serum ferritin was found a good predictor for predicting babies with IUGR. The data above depict that women with mean serum ferritin above 18.2 ng/ml, were 10.23 times more likely to get asymmetrically growth restricted babies (CI 2.89–36.17, $p<0.001$).

Discussion

Fetal growth restriction is a serious condition deserving early diagnosis or good predictive tests to help reduce the impact of its serious short and long term complications. The current study found a significantly higher mean maternal serum ferritin level in the group of pregnant women who ended up being diagnosed with IUGR, as opposed to that in control pregnancies (19.3 ± 6.83 vs. 14 ± 5.18 mg/L, respectively; $p<0.001$). This is, roughly, similar to the so far recorded ferritin levels in similar studies. The value of maternal serum ferritin in predicting asymmetric IUGR was previously investigated, for few times, on small numbers of cases, i.e. 7 cases in Ozgu-Erdinc et al (9), 18 cases in Milasinovic et al (13), 10 cases in Hubel et al (14), 17 cases in Visnjevac et al (15), and 31 cases in Bindal et al (16), or with limitations in the research methodology such as measuring ferritin at a gestational age that was too early (20 weeks) or too late (36 weeks) (1,4), or including cases with maternal iron deficiency anemia (15).

Lower expression of placental transferrin is known to be associated with IUGR (18). This may cause less iron extraction by the placenta from maternal serum, thus raising the level of serum fer-

ritin (16). Furthermore, other studies suggest that placental damage, in cases of asymmetric IUGR, releases some of the placental ferritin into the maternal circulation, thus further raising its serum level (1,14,19). Ferritin is also a known acute phase reactant released by leukocytes into the systemic circulation in response to immune activation. High serum ferritin level may reflect a non-infectious vascular inflammatory response causing oxidative damage to cells and tissues (14,20). The association of IUGR with inflammation has been previously proven (21). This is the reason why this study set acute infections during pregnancy, leukocytosis or raised C-reactive protein titer among its exclusion criteria to rule out any misleading rise in the level of ferritin.

As opposed to previous studies (3,15,16), we set anemia among our exclusion criteria. Our results showed non-significant differences between controls and IUGR cases as regards erythrocytic counts, hematocrit and hemoglobin levels, compared to a rate of anemia of 47.4% in the control group of one study (15). This is probably why the current study found a higher degree of correlation between maternal serum ferritin and neonatal birthweight (-0.453) which is higher than those found by Višnjevac et al (-0.24) (15), and Bindal et al (-0.36) (16). On other grounds, Akkurt et al reported a higher hematocrit level in pregnancies with IUGR than controls (1). They hypothesized that this was related to a degree of hemoconcentration related to IUGR development. The current study did not show such difference in hematocrit.

We showed maternal serum ferritin to have a good predictive power when used at 30-32 weeks for prediction of IUGR. The results of the current study suggest that a ferritin level of >18.2ng/mL can predict asymmetric IUGR with a sensitivity of 59.38%, a specificity of 90.62%, a positive predictive value of 86.4%, and a negative predictive value of 69%. The very good overall accuracy of 76.8% drawn from the ROC analysis and the associated odds ratio of 10.23 are promising, denoting that serum ferritin measurement may be considered an adequate predictive test for IUGR. Uberos et al, Višnjevac et al and Bindal et al also came to similar conclusions but with a lower odds ratio

(3,15,16), probably due to the inclusion of cases with anemia in the study population (3,16). Uberos et al concluded that ferritin levels >13 ng/ml had 4.5 times more risk to result in IUGR at 38 weeks of gestation (3). Hubel et al showed no significant difference in ferritin between AGA and IUGR cases, but their study only included 10 asymmetric IUGR cases (14). Soubasi et al found high maternal ferritin levels to correlate significantly with higher rates of gestational diabetes mellitus and IUGR, but did not define a cutoff to differentiate IUGR from AGA neonates (22). Hou et al, Milasinovic et al and Višnjevac et al identified the ferritin threshold to predict IUGR or SGA as >13.6 ng/ml (4,13,15). Ozgu et al reported maternal serum ferritin levels >60 ng/ml to increase the risk for IUGR (9). Bindal et al concluded that maternal serum ferritin >20 ng/ml predicted IUGR with an odds ratio of 6.6 (16). Akkurt et al took one step further and showed a role for maternal serum ferritin measurements in distinguishing pregnancies with asymmetric IUGR from those with simple small for gestational age babies and no signs of placental insufficiency (1).

In conclusion, this study presents further evidence, on a relatively larger group of patients, that maternal serum ferritin might serve as a useful marker to identify pregnancies at risk of resulting in asymmetric IUGR infants. It found a significantly higher maternal serum ferritin level, at 30-32 weeks, in pregnancies destined to develop asymmetric IUGR at a later gestational age than in controls. A cutoff of >18.2 ng/mL had an accuracy of 76.8% to predict IUGR with a PPV of 86.4%. Pregnancies with ferritin level higher than such cutoff were 10.23 times more likely to deliver asymmetrically growth restricted babies when compared to women with serum ferritin value less than <18.2 ng/ml. Large scale studies are recommended to further establish this theory and to test related points regarding the effect of associated anemia or the gestational age at which the test is best to be done. The role of maternal serum ferritin in distinguishing pregnancies with asymmetric IUGR and signs of placental insufficiency from those with simple small for gestational age babies also still needs further research.

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Conflicts of interest

The authors have stated explicitly that there are no conflicts of interest in connection with this article.

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Figures Legends:

Figure (1) Receiver operating characteristic curve showing the best cutoff point for serum ferritin level between mothers with asymmetric IUGR neonates and those with AGA, was >18.2 ng/mL.

Tables Legends:

Table (1) Comparison between cases and controls regarding laboratory and Doppler variables.

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Table (3) Correlation between serum ferritin level and maternal and neonatal data in cases and controls.

Table (4) Correlation between the development of IUGR in the current pregnancy and the past history of previous IUGR deliveries, current pregnancy serum ferritin level or umbilical and middle cerebral artery Doppler indices.

Table (5) Data showing sensitivity, specificity, positive predictive value, negative predictive value of various serum cutoffs to predict asymmetrical IUGR.

Table (1) Comparison between cases and controls regarding laboratory and Doppler variables.

		Cases group n=32	Control group n=32	p value
Erythrocytes (10 ⁶ /μl)	Mean (±SD)	4.12 (±0.58)	3.88 (±0.66)	0.13
	Range	3.2 – 5.23	3.04 – 5.04	
Hemoglobin (gm/dl)	Mean (±SD)	11.5 (±0.43)	11.4 (±0.36)	0.12
	Range	11 – 12.5	11 – 12.3	
Hematocrit (%)	Mean (±SD)	34.6 (±1.84)	33.9 (±1.59)	0.12
	Range	31.3 – 38.6	30.8 – 37.8	
Leucocytes (10 ³ /cc)	Mean (±SD)	8.6 (±2.13)	8.5 (±1.76)	0.84
	Range	5.4 – 12.3	5.4 – 12.2	
Ferritin (ng/ml)	Mean (±SD)	19.3 (±6.83)	14 (±5.18)	<0.01
	Range	8.5 – 40	5 – 26.3	
Umbilical artery Doppler	RI	Mean (±SD)	0.71 (±0.06)	<0.01
		Range	0.6 – 0.81	
	PI	Mean (±SD)	1.24 (±0.17)	<0.01
		Range	1.03 – 1.94	
Middle cerebral artery Doppler	RI	Mean (±SD)	0.74 (±0.09)	<0.01
		Range	0.61 – 0.92	
	PI	Mean (±SD)	1.64 (±0.07)	<0.01
		Range	1.36 – 1.88	

N=number of women, SD=standard deviation, RI=resistance index, PI=pulsatility index.

Table (2) Comparison between cases and controls regarding neonatal outcome at delivery.

		Cases group n=32	Control group n=32	p value
Gestational age at delivery (days)	Mean (\pm SD)	272.9 (\pm 7.25)	274.3 (\pm 6.79)	0.14
	Range	259 – 282	260 – 285	
Mode of delivery (n, %)	Cesarean	19, 59.4%	16, 50%	0.45
	Vaginal	13, 40.6%	16, 50%	
Birth Weight (gm)	Mean (\pm SD)	2134 (\pm 143)	3419 (\pm 352)	<0.001
	Range	1750 – 2390	2800 – 4100	
Apgar score (1st minute)	Median (IQR)	7 (7–8)	8 (7–9)	0.04
	Range	6 – 9	6 – 10	
Apgar score (5th minute)	Median (IQR)	8.5 (8–9.5)	9 (9–10)	0.03
	Range	7 – 10	8 – 10	
Admission to NICU (n, %)	No	19, 59.4%	28, 87.5%	0.01
	Yes	13, 40.6%	4, 12.5%	

N=number of women, SD=standard deviation, IQR=interquartile range, NICU=neonatal intensive care unit.

Table (3) Correlation between serum ferritin level and maternal and neonatal data in cases and controls.

	Ferritin level (ng/ml)			
	Cases		Controls	
	r value	p value	r value	p value
Maternal Age (years)	-0.195	0.28	-0.112	0.54
Erythrocytes ($10^6/\mu$ l)	0.111	0.55	-0.073	0.69
Hemoglobin (gm/dl)	0.080	0.66	0.148	0.42
Hematocrit (%)	0.225	0.22	0.044	0.81
Leucocytes ($10^3/$ cc)	0.250	0.17	-0.109	0.56
Gestational age at delivery (days)	-0.261	0.15	0.103	0.57
Birth Weight (gm)	-0.453	0.009	0.174	0.34
Apgar score (1st minute)	-0.268	0.14	0.063	0.73
Apgar score (5th minute)	-0.090	0.63	0.083	0.65

r value = Spearman coefficient of correlation.

Table (4) Correlation between the development of IUGR in the current pregnancy and the past history of previous IUGR deliveries, current pregnancy serum ferritin level or umbilical and middle cerebral artery Doppler indices.

	Development of IUGR in the current pregnancy	
	r value	p value
Previous IUGR	1.435	0.09
Ferritin (ng/ml)	0.183	0.001
Umbilical artery RI	16.866	<0.001
Umbilical artery PI	6.596	<0.001
Middle cerebral artery RI	-12.308	0.002
Middle cerebral artery PI	-17.688	0.03

r value = Spearman coefficient of correlation.
 RI=resistance index, PI=pulsatility index.

Table (5) Data showing sensitivity, specificity, positive predictive value, negative predictive value of various serum cutoffs to predict asymmetrical IUGR.

Cutoff point	AUC	Sensitivity	Specificity	PPV	NPV
>14.7	0.734	75.00	71.87	72.7	74.2
>15.6	0.687	65.62	71.87	70.0	67.6
>16.4	0.719	65.62	78.12	75.0	69.4
>17	0.703	59.38	81.25	76.0	66.7
>18.2	0.768	59.38	90.62	86.4	69.0
>20.1	0.625	34.38	90.62	78.6	58.0
>22	0.641	34.38	93.75	84.6	58.8
>25	0.516	9.38	93.75	60.0	50.8
>26	0.531	9.38	96.87	75.0	51.7

AUC=area under the curve, PPV=positive predictive value, NPV=negative predictive value.

Figure (1)
 Receiver operating characteristic curve showing the best cutoff point for serum ferritin level between mothers with asymmetric IUGR neonates and those with AGA, was >18.2 ng/mL.

